Japan Customs Analysis Methods

No. 302

Analysis Method of Peppermint Oil

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1. Scope

This analysis method is applied to peppermint oils of subheadings 3301.24 and 3301.25 of Customs Tariff Law (Appendix Table – Customs Tariff Schedule), for distinguishing between those obtained from Mentha piperita and those from M. arvensis.

2. Outline of Test Method

Under Customs Tariff Law (Appendix Table – Customs Tariff Schedule), peppermint oils are divided into three groups, i.e. those obtained from M. piperita, those obtained from M. arvensis and others. This analysis method enables to distinguish between peppermint oil obtained from M. piperita and that from M. arvensis.

The peppermint oils from these two Mentha species are distinguished using gas chromatograph mass spectrometer, by detecting menthofuran, sabinene hydrate and viridiflorol, all of which are specifically contained in those obtained from M. piperita, and comparing the content ratio of cineole and limonene commonly contained in these two types of oils.

3. Apparatus

(1) Gas chromatograph-mass spectrometer (GC-MS)
(2) Capillary column
  Use a DB-WAX (30 m × 0.25 mm i.d., film thickness 0.5 µm) or equivalent one
(3) GC-MS, operating under the following conditions:
  (a) GC temperature program
      Program the oven temperature so that each constituents of peppermint oils are separated sufficiently (e.g., initial temperature 50 °C for 3 min; increase at 15 °C per min; 110 °C; increase at 3 °C per min; 150 °C; increase at 15 °C per min; final temperature 200 °C for 5 min).
  (b) Injection port temperature · 250 °C
  (c) Others
      Set other instrument parameters to the optimum conditions for the analysis.

4. Reagents

(a) Standard peppermint oil
  Use peppermint oils obtained from M. piperita and M. arvensis whose constituents being main peaks in their gas chromatograms have been identified.
(b) Diethyl ether
    JIS special reagent grade or equivalent

Note 1) Standard peppermint oils have to be stored in a cool and dark place.

5. Procedure

5.1. Measurement

Prepare a test solution of approx. 1 % concentration by adding diethyl ether to the test sample. Inject 1 μl of the test solution into GC-MS to obtain a total ion chromatogram and to measure mass spectra of object peaks.

5.2. Identification of constituents

Identify object peaks in the total ion chromatogram of the test sample, based on their mass spectra and by comparing their retention times with those of the same compounds present in the standard peppermint oils.
Note 2) When additional peaks other than peaks in the total ion chromatograms of the standard peppermint oils are found, the presence of additives in the test sample should be confirmed.

Measurement conditions: Column, DB-WAX (30 m × 0.25 mm i.d., film thickness of 0.5 µm); Oven temperature, initial 50 °C (3 min.) □ 15 °C/min → 110 °C → 3 °C/min → 150 °C → 15 °C/min → final 200 °C (5 min); Carrier gas, Helium (1 ml/min); Injected volume, 1 µl; Injection port temperature, 250 °C; Injection port mode, sprit (20:1); Interface temperature, 250 °C; Ionization, EI, 70 eV.

Reference 1: Examples of total ion chromatograms

<table>
<thead>
<tr>
<th>Peak No.</th>
<th>Compound</th>
<th>Peak No.</th>
<th>Compound</th>
<th>Peak No.</th>
<th>Compound</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>α-Pinene</td>
<td>6</td>
<td>(+) trans-Sabinene hydrate</td>
<td>11</td>
<td>l-Menthol</td>
</tr>
<tr>
<td>2</td>
<td>β-Pinene</td>
<td>7</td>
<td>Menthone</td>
<td>12</td>
<td>Piperitone</td>
</tr>
<tr>
<td>3</td>
<td>d-Limonene</td>
<td>8</td>
<td>Menthofuran</td>
<td>13</td>
<td>Viridiflorol</td>
</tr>
<tr>
<td>4</td>
<td>Cineole</td>
<td>9</td>
<td>iso-Menthone</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>3-Octanol</td>
<td>10</td>
<td>Menthylacetate</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
5.3. Identification of peppermint oils

Identify the kind of peppermint oil sample using the table below indicating differences between peppermint oil obtained from *M. piperita* and that obtained from *M. arvensis*.

If the test samples are mixtures of the two kinds or obtained from hybrids between the two species, it is difficult to identify them using the criteria of the table below. In that case, comprehensive analysis is required, including quantitative studies on their physiochemical properties (specific gravity, refractive index, optical rotation and the like) and chemical compositions.
Table  Criteria for discrimination between two kinds of peppermint oils

<table>
<thead>
<tr>
<th>Item</th>
<th>Kind of peppermint oil</th>
<th>M. piperita</th>
<th>M. arvensis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Detection of menthofuran</td>
<td>(+)</td>
<td></td>
<td>(-) (3)</td>
</tr>
<tr>
<td>Detection of sabinene hydrate</td>
<td>(+)</td>
<td></td>
<td>(-) (3)</td>
</tr>
<tr>
<td>Detection of viridiflorol</td>
<td>(+)</td>
<td></td>
<td>(-) (3)</td>
</tr>
<tr>
<td>Peak height of limonene and cineol</td>
<td>limonene &lt; cineol</td>
<td>limonene &gt; cineol</td>
<td></td>
</tr>
</tbody>
</table>

Note 3: Where, "(-)" indicates that the compound concerned is undetectable at a standard sensitivity level (which allows detection of the compound from standard M. piperita but not from M. arvensis).

5.4. Determination of Total Menthol

If the sample is identified to be peppermint oil obtained from M. arvensis according to the procedure in 5.3, its total content of menthol is determined by the following quantitative method based on the “Mentha oil” documented in the Japanese Pharmacopoeia (11th Edition).

Weigh about 10 g of the test sample into an extraction flask. Add 10 ml of acetic anhydride and 2 g of sodium acetate, anhydrous to the flask. Attach a reflux condenser to the flask and boil for 1 hour. Cool it and add 30 ml of water. Heat the flask in boiling water while stirring occasionally for 15 min.

Transfer the contents to a separating funnel to separate the oil layer. Wash the oil layer with water until it becomes neutral. Add 2 g of sodium sulfate, anhydrous, and shake it vigorously. After leaving to stand for 30 min, filter it through a dried paper filter.

Accurately weigh about 2 g of the obtained (acetylated) oil and about 5 g of the test sample (unacetylated oil) into separate extraction flasks. Add 5 ml of ethanol and two drops of phenolphthalein solution (4) to both flasks. After neutralizing both solutions by adding 0.5 mol/l of potassium hydroxide-ethanol solution, add 25 ml of 0.5 mol/l potassium hydroxide-ethanol solution. Attach a reflux condenser to the flask and boil the content for 1 hour. Add 25 ml of water immediately after boiling and stir. Cool the solution and titrate the excess amount of potassium hydrate with 0.5 mol/l of hydrochloric acid. Perform a blank test in the same manner.

\[
7.814 \times (c-a) \left( \frac{1 - \frac{0.021 \times (c-b)}{[\text{unacetylated oil (g)]}}} {[\text{acetylated oil (g)] - 0.021 \times (c-a)}} \right)
\]

Where: “a” and “b” represent the titration volumes (ml) of 0.5 mol/l of hydrochloric acid added to acetylated oil and unacetylated oil, respectively. “c” represents the titration volume (ml) of hydrochloric acid used in the blank test.

Note 4: Phenolphthalein solution is prepared by dissolving 1 g of phenolphthalein in 100 ml of ethanol.

6. References

1. Tada et al., (1971) Reports of the Central Customs Laboratory, 5, 59-64
2. Formacek and Kubeczka: Essential Oil Analysis, John Wiley & Sons
4. 梅本和泰：ハッカ属植物の自殖株の精油成分 (Japanese only)